

FUJIFILM



IrvineScientific

## CHANG Marrow Bone Marrow Culture Medium with Gentamicin

Catalog No. 91031

100mL, 500mL

For *in vitro* diagnostic use.Zur *In-vitro*-Diagnostik.Solo per uso diagnostico *in vitro*.Para uso diagnóstico *in vitro*.Pour diagnostics *in vitro*.Para utilização em diagnóstico *in vitro*.Για *in vitro* διαγνωστική χρήση.Pro diagnostické použití *in vitro*.Til *in vitro*-diagnostik.*In vitro* -diagnostikkaan.Lietošanai *in vitro* diagnostikā.Uitsluitend voor *in vitro* diagnostisch gebruik.Do diagnostyki *in vitro*.Pentru uz diagnostic *in vitro*.För *in vitro*-diagnostik.*In vitro* diagnostiliseks kasutamiseks.*In vitro* diagnosztikai alkalmazáshoz.Skirta *in vitro* diagnostikai.*In vitro* diagnostik kullanim için.Na diagnostické použitie *in vitro*.За *in vitro* диагностична употреба.За употребу *in vitro* dijagnostici.Ghal úžu dijanjostiku *in vitro*.За diagnostično uporabo *in vitro*.

### REFERENCES

Tijo, JH, and Whang-Peng, J: Direct Preparation of Bone Marrow Cells. Human Chromosome Methodology (JJ Yunis, ed.), Academic Press, New York, 1974.

Hozier, JC, and Lindquist, L: Banded Karyotypes from Bone Marrow: A Clinically Useful Approach. Human Genetics, 53:205-209, 1980.

Williams, DL, et al: A Direct Bone Marrow Chromosome Technique for Acute Lymphoblastic Leukemia. Cancer Genetics and Cytogenetics, 13:239-257, 1984.

Babior, B, and Stossel, T: Hematology, A Pathophysiological Approach, Churchill-Livingstone, Inc., New York 1990.

LeBeau, M: Cytogenetic Analysis of Hematologic Malignant Diseases. ACT Cytogenetics Laboratory Manual, Raven Press, New York, 1991.

Mitelman, F.: Catalog of Chromosome Aberrations in Cancer (4<sup>th</sup> ed.), Alan Liss, New York, 1991.

Kaplan, B, and Dale, K (eds): The ACT Cytogenetic Symposia, CA 1994.

Mitelman, F, and Heim, S: Cancer Cytogenetics, Wiley-Liss, New York, 1995.

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### Glossary of Symbols\*:

REF

Catalog Number

LOT

Lot Number

STERILE A

Sterilized using aseptic processing techniques (filtration)

Expiration:  
Year - Month - Day

Caution, consult accompanying documents



Consult instructions for use

Storage Temperature  
store below -10°C

Do not resterilize



Do not use if package is damaged



Manufacturer



CE Mark

EC REP

Emergo Europe - Prinsessegracht 20  
2514 AP The Hague  
The Netherlands

\*Symbol Reference - EN ISO 15223-1, Medical devices – Symbols to be used with medical device labels, labeling.

### INDICATION FOR USE

CHANG Marrow is intended for use in primary culture of clinical Human Bone Marrow Cultures for karyotyping and other genetic testing of various hematological disorders.

### DEVICE DESCRIPTION

CHANG Marrow is a ready-to-use medium consisting of IMDM, with FBS, HEPES buffer, L-glutamine, Giant Cell Tumor (GCT) Conditioned Medium, Recombinant Human GM-CSF and Gentamicin Sulfate. CHANG Marrow has been optimized to support efficient growth of bone marrow cells for cytogenetic analysis. No addition of any components prior to culturing bone marrow is required. CHANG Marrow contains Gentamicin Sulfate (50 mg/L). Additional antibiotics may be added if desired.

### COMPONENTS

Amino Acids	Proteins, Hormones, and Growth Factors	Energy Substrates
Alanine		Glucose
Arginine		Pyruvate
Asparagine		Inositol
Aspartic Acid	Fetal bovine serum (FBS)	Antibiotic
Cystine	hrGM-CSF	Gentamicin Sulfate
Glutamic Acid		
Glutamine		
Glycine	Salts & Ions	Other
Histidine	Sodium chloride	Biotin
Isoleucine	Sodium selenite	Giant cell tumor conditioned medium (GCT-CM)
Leucine	Calcium chloride	
Lysine	Choline chloride	
Methionine	Potassium chloride	Vitamins and trace elements
Phenylalanine	Potassium nitrate	Folic acid
Proline	Magnesium sulfate	Nicotinamide
Serine	Sodium phosphate	Riboflavin
Threonine		Thiamine
Tryptophan	Buffers	Pantothenic acid
Tyrosine	Sodium bicarbonate	Cobalamin
Valine	HEPES	Pyridoxine
		Water
		WFI Quality

### QUALITY ASSURANCE

Several factors including source of specimens, culture conditions and selection of reagents can influence the result obtained. Users are advised to run each new batch of reagent in parallel with reference material of known suitable activity before adoption in routine use. Each lot of CHANG Marrow has been performance tested on Clinical Bone Marrow Cultures at an independent Clinical Cytogenetics Laboratory compared to a control medium. Results are reported on a lot specific Certificate of Analysis.

### MATERIALS AND EQUIPMENT REQUIRED BUT NOT PROVIDED

1. Plastic Sterile Centrifuge Tubes and Culture Flasks
2. CO<sub>2</sub> Incubator at 37°C
3. Bench Centrifuge
4. Vortex Mixer
5. Colcemid Stock Solution, 10 µg/mL
6. Potassium Chloride Solution, 0.075 M
7. Fixative Solution, Methanol:Acetic Acid (3:1)

### PREPARATION FOR USE

Thaw overnight in the refrigerator (2 - 8°C), then gently mix to ensure homogeneity. Aseptically dispense 10 mL of medium into sterile culture flasks and equilibrate to 37°C for immediate use.

### DIRECTIONS FOR USE

#### I. Sample Preparation:

Use 0.5 to 1.0 mL of sodium heparinized bone marrow aspirate. Lithium heparin, EDTA, or citrate anticoagulants are unsuitable for cytogenetic studies.

- If more than 5 mL of bone marrow aspirate is received, the sample may be hemodilute. Spin the specimen down at 1,200 rpm for 8 minutes to isolate the bone marrow fraction.

- If the specimen arrives in transport medium, spin the sample down at 1,200 rpm for 8 minutes and remove the transport medium (supernatant). Inoculate using the remaining spun-down fraction in the bottom of the tube.

For additional details on the use of these products, each laboratory should consult its own laboratory procedures and protocols which have been specifically developed and optimized for your individual medical program.

#### II. Bone Marrow Culture:

Label all culture vessels with patient name, specimen number, and culture type.

1. Before inoculation of specimen bring CHANG Marrow to ambient temperature.
2. Inoculate each culture with the appropriate amount of sample to achieve an optimal concentration of 1x10<sup>6</sup> cells/mL or 10 x 10<sup>6</sup> cells per 10 mL culture.
3. Each individual laboratory should determine the number of cultures to set up depending on the clinical indication of the patient. Additional growth factors may be added if desired.
4. Incubate cultures at 35 - 39°C, 5-8% CO<sub>2</sub> atmosphere until ready for harvest.

#### Harvesting the Cultures:

1. Remove the culture ready for harvest from the incubator and gently swirl flask to re-suspend cells.
2. Transfer the contents of each flask to a 15 mL centrifuge tube.
3. Add 40 µL of stock Colcemid (10µg/mL) to each culture tube. Tightly cap tubes and mix gently by inverting.
4. Incubate tubes at 35-39°C, for 45 minutes.
5. After incubation, centrifuge tubes for 8 minutes at 1,000 rpm.
6. Carefully aspirate supernatant from each tube using a vacuum aspirator, with solvent trap. Be careful to not aspirate pellet.
7. Resuspend cell pellet by tapping bottom or side of each tube with finger.
8. Initiate a 20 minute timer.
9. Add 3 - 4 mL dropwise, of prewarmed (35 - 37°C) hypotonic solution (0.075M Potassium Chloride).
10. Tightly cap tube and mix gently by tapping bottom or side of the tube with finger.
11. Add 5 - 6 mL dropwise, of prewarmed (35 - 37°C) hypotonic solution. Tightly cap tube and invert tube.
12. Repeat Steps 9 - 11 for each tube.
13. Using a water bath, allow tubes to stand at 35 - 37°C. Invert tubes once at midpoint of 20 minute timer.
14. At the end of the 20 minute timer, remove tubes from water bath and add 1 mL of fresh 3:1 Carnoy's Fixative to each tube. Tightly cap and invert each tube. (This is the Pre-Fixative step)
15. Centrifuge tubes for 8 minutes at 1,000 rpm.
16. Aspirate supernatant from each tube, leaving about 1 mL above the cell pellet. Be careful to not aspirate pellet. Be cautious of fibrous material that may extend from the cell pellet up into the supernatant after centrifugation. The last few mL of supernatant may need to be removed by hand with a Pasteur pipette (not using vacuum aspiration) to avoid aspirating the entire cell pellet into the waste container.
17. Resuspend cell pellet, as described in Step 7.
18. Add 3 - 4 mL dropwise of fresh 3:1 Carnoy's Fixative.
19. Add remaining fixative up to 7 mL.
20. Repeat steps 16 - 19 for each tube.
21. Let stand for 10 minutes at room temperature. (This is the First fixative step).
22. Centrifuge tubes for 8 minutes at 1,000 rpm.

23. Aspirate supernatant leaving about 1 mL above pellet. Resuspend cell pellet.
24. Add fix, up to 7 mL. Centrifuge tubes for 8 minutes at 1,000 rpm. (Second fixative step).
25. Repeat steps 22 - 23. (Third fixative step).
26. At this point, fixed cells pellets can be used immediately for slide preparation according to the laboratory standard protocol or stored in the refrigerator (2 - 8°C) or freezer for future use.

### STORAGE AND STABILITY

CHANG Marrow should be stored frozen below -10°C until ready to use. CHANG Marrow is stable until the expiration date shown on the bottle label when stored frozen. After thawing, any unused product can be dispensed into working aliquots and refrozen for later use, or tightly capped and stored at 2 - 8°C for up to 30 days. Protect from fluorescent light.

### PRECAUTIONS AND WARNINGS

This device is intended to be used by staff trained in procedures that include the indicated application for which the device is intended.

CHANG Marrow contains FBS and GCT conditioned medium and should be handled with universal laboratory precautions. The medium contains an antibiotic (gentamicin sulfate) to reduce the potential of bacterial contamination, but aseptic techniques should always be used when dispensing the medium. Do not use any medium that is not red in color.









