

# TIPS FOR HANDLING ABNORMAL SPERM

**PREPARING SPERM FOR IUI, IVF, OR ICSI CAN ENCOUNTER A VARIETY OF COMPLICATIONS. IRVINE SCIENTIFIC HAS COMPILED A SERIES OF TIPS FROM DR. JAYANT G MEHTA FROM THE QUEEN'S HOSPITAL IN ROMFORD ESSEX TO HELP DEAL WITH ABNORMAL SPERM SAMPLES. FOR GENERAL SPERM SEPARATION AND PROCESSING, PLEASE REFER TO OUR PRODUCT INSERTS OR ANDROLOGY PROTOCOLS.**

*Irvine Scientific has not validated these procedures and each laboratory should consult its own laboratory procedures and protocols which have been specifically developed and optimized for your individual medical program.*

The following tips feature the use of ISolate® (Catalog # 99264) for gradient sperm separation and Multipurpose Handling Medium® (MHM®, Catalog # 90163) for sperm washing and handling. For different gradient percentages of ISolate, please dilute with MHM following the directions found in the product insert. Dr. Mehta uses a 40%/80% ISolate gradient for normal sperm processing.

## **SPECIMEN ISSUE:**

### **Low sperm concentration and/or reduced progressive motility of <30%**

- Use 1 mL layers of 30% and 60% ISolate gradient.
- Resuspend the resulting pellet in 0.5 mL of MHM.
- Follow with swim up by layering 0.5 mL of MHM.
- Incubate for 30-45 minutes in a heating block (not CO<sub>2</sub> incubator).
- Transfer the top 40% of media to a new tube and count motile sperm.
- Remove a further 20% if few or no sperm are seen.

## **SPECIMEN ISSUE:**

### **High cellular debris**

- To prevent debris from clogging the gradient interfaces, load ISolate with small volumes of semen sample at <0.5 mL.
- Instead of layering the semen sample, gently mix the semen sample directly into the top 10% of the ISolate upper layer.
- Due to the smaller volumes, this will require multiple gradient tubes.

## **SPECIMEN ISSUE:**

### **Retrograde ejaculated urine specimens**

- Centrifuge urine at 1600 RPM for 10 minutes.
- Remove supernatant and dilute the pellet by adding 3mL of MHM.
- Layer the resuspended pellet on 40% ISolate gradient.

## **SPECIMEN ISSUE:**

### **High Viscosity**

#### **METHOD 1:**

- Mix the semen sample with equal volume of MHM prior to gradient layering.
- If sample fails to disperse in 2-3 minutes, place it in 37°C for 10-15 minutes.
- Samples displaying high viscosity may require gentle repeated pipetting or passage through a blunt 18 gauge needle as per the WHO manual. Use of blunt needle avoids damaging sperm during pipetting. Alternatively, you can use a sterile glass pipette to do the same. Pipetting 8-10 times normally breaks down the viscous sample.

#### **METHOD 2:**

- Use 0.2% solution of a-amylase for 5 minutes.
- Wash the enzyme on liquefaction: after 5 minutes, add 5 mL of MHM and centrifuge at 1600 RPM for 5 minutes.
- Remove the supernatant and resuspend the pellet in 1 mL of MHM.
- Layer the resuspended pellet on 40% ISolate.

## **SPECIMEN ISSUE:**

### **Cryopreserved semen sample**

- Since osmolality of semen cryoprotectant is high, the spermatozoa will swell greatly upon entering the 40% ISolate layer and becoming too buoyant.
- Slowly dilute the thawed cryoprotectant semen with 5X volume of MHM, adding drop-wise with constant gentle mixing over a period of 15 minutes.
- Layer on 40% ISolate gradient.

\*Dr. Jayant G Mehta is one of the editors of Male Infertility – Sperm Diagnosis, Management and Delivery.



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